Microbial influence on the sorption of ¹³⁷Cs onto materials relevant to the geological disposal of radioactive waste

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Summary. A major concern in the geological containment of radioactive wastes is the speed of movement of radionuclides from the repository, after their eventual leaching and release, into the geosphere and finally into the biosphere. Radionuclide sorption onto the host rock is an important retarding mechanism. Experimental evidence shows that the presence of microbes in this environment influences the sorption capabilities of the host rock. Their presence can decrease the amount of retardation of ¹³⁷Cs, a common radionuclide in radioactive waste, by the solid phase. Sorption methods and data analysis procedures are presented and the implications for radioactive waste disposal assessments are discussed.

Key words. Microbiology; radioactive waste; geology; radionuclide sorption; ¹³⁷Cs.

Introduction

Radioactive waste can be gaseous, liquid or solid and arises in many different physical forms and concentrates. The safe disposal of these wastes is a major technical issue. Many countries favour disposal of solid wastes in repositories in geological formations to ensure that the waste is isolated from man's environment and from major natural processes or intrusion by man and to provide a shield from the radiation^{2,3}. The processes which will affect containment and behaviour of the waste are dominated by the behaviour of groundwater in the host rock and other rock formations above and below a repository. The movement of groundwater through a repository will result in leaching and mobilisation of radionuclides ^{2,3,12,17}.

It is now assumed 4, 22, 23 that microbes exist in geological formations relevant to the disposal of radioactive waste and that some groups can tolerate the extreme environmental conditions present³. Microbes are implicated in biodeterioration of materials that will be used in repository construction and cause geochemical changes in some environments. Microbes mobilise U13 and take up metals 9, 16, 21, 27, thus they could have a significant role in the mobilisation and transport of radionuclides. The processes involved in radionuclide migration from a repository are complex even without the possible influence of microorganisms. Following failure of the engineered barriers in a repository, retardation of movement of remaining radionuclides will depend on the properties of the host rock. The time to reach the biosphere and the concentrations in which they will arrive are related to the distance and velocity of their movement through the host rock and depend on the geochemistry and physical properties along the migration path³.

Chapman and McKinley³ define sorption mechanisms

- as including purely physical processes retarding migration such as molecular filtration, ion exclusion and diffusion into dead-end pores,
- direct chemical reaction with rock surfaces involving physical adsorption, chemical adsorption or direct incorporation into the rock structure,

- and indirect chemical reactions such as precipitation caused by enhanced concentrations at the rock surface. 'Sorption' mechanisms can be measured statically, using batch studies and measuring the partitioning of a radionuclide spike between the solid and liquid phases or dynamically using column experiments where a radionuclide spike is passed through a rock/soil column and the concentration profile in the effluent or the profile in the column is used to give a retardation factor ³.

The influence of microorganisms on such sorption experiments will also give some indication of the significance of microorganisms in the far field. In this paper the effect of sulphate reducing bacterial (SRB) on sorption of ¹³⁷Cs onto Fullers' Earth (calcium montmorillonite), a potential backfill material, is monitored using a traditional batch sorption method with distribution ratios (Rd) being produced. Sulphate reducing bacteria are of particular interest in waste disposal because they are important in anaerobic corrosion processes ¹⁹; their activity can change the geochemical environment ²⁸; and their tolerance to repository conditions has been established ^{24, 25}.

Materials and methods

The batch sorption method used by West et al. ²⁵ was applied to a study of radiocaesium sorption on Fullers' Earth with an isolate of SRB (*Desulfovibrio desulfuricans*) from Konrad mine (KSRB), West Germany ⁴. The methodology consists of contacting the pure rock, and a mixture of rock and microorganisms, with groundwater (filter sterilised through a 0.2- μ m filter) spiked with ¹³⁷Cs under a variety of conditions. Solution to solid ratios remained constant at 10:1 ml/g. Systems were incubated under anaerobic or aerobic atmospheres at room temperature. Five concentrations of ¹³⁷Cs from 7.3 × 10⁶ to 7.3 × 10¹⁰ M (1 to 10⁻⁴ ppm) were prepared from standards (Amersham). All tests were run in duplicate and for two equilibration periods (1 and 8 days).

Microbial cultures had been grown in groundwater from the Fullers' Earth quarry in anaerobic conditions for 1 and 1-4 days prior to use in the experiment. An inherent difficulty is the inevitable variation in microbial numbers and the fact that sorption could occur onto actively growing, dormant as well as dead microorganisms. Solid and liquid phases were separated by centrifugation followed by filtration (0.2 µm pore size) since centrifugation alone did not adequately separate the phases. The dry weight of the microbes was ascertained by weighing dried filters before and after filtration which would include the mass of all the organisms (in all conditions) plus any associated organic or inorganic material. Unfortunately this method does not trap particles below 0.2 µm as ultra filtration would have done. Microbial numbers at the beginning and end of the experiment were determined using a standard spread plate technique on Postgates Medium E 19,26 . Approximately 4×10^6 SRB were introduced into each test from 1-day-old cultures and approximately 3×10^4 from 14-day-old cultures. Numbers were assumed to be unchanged after 1 day's equilibration. No SRB survived after 8 days aerobic equilibration. Anaerobic SRB growth in the groundwater medium had already been established. Figures were not obtained for growth after 8 days.

Samples of rock and groundwater were collected from Baulking Quarry, Fernham, Oxfordshire, England (Brett Bentonite Co). The detailed geology of the area is described by Poole and Kelk ¹⁸. The rock phase used in these experiments was Fullers' Earth (calcium montmorillonite) whose microbial content has already been ascertained. The material was characterised using XRD and grain size analysis ²⁶. It was treated in three ways: wet sterilised in an autoclave; dry sterilised in a warm air oven; and left unaltered. Sorption was corrected for the variable moisture remaining. Analysis of the groundwater ²⁶ shows high concentrations of Na and Ca, as a result of interaction with the clay via ion exchange. Cs was absent in the groundwater (> 0.05 ppb) and sulphate was the predominant anion.

The kinetics of sorption of ¹³⁷Cs onto Fuller's Earth was complete after 24 h.

After the allocated equilibration time, the extent of radionuclide sorption was evaluated by measuring the change in 137 Cs activity in the aqueous phase by gamma spectrometry using a shielded NaI Gamma detector. The distribution ratios (Rd) were ascertained using the equation described by West et al.²⁵ and represent the ratio of sorbed to dissolved 137 Cs, i.e., Rd = C_R/C_W , where C_R is the concentration in the solid phase (g g⁻¹) and C_W is the concentration in the aqueous phase (g ml⁻¹).

Results

Summaries of the calculated distribution ratios (Rd) are given in tables 1-4, the full data sets are presented in West et al.²⁶.

Table 1. Rd values for aerobic conditions with rock only (ml g⁻¹)

	, , ,	
1 day equilibration	8 days equilibration	
157	194	
195	220	
223	294	
193	266	
232	273	
558 (441)	537 (423)	
682 (539)	788 (622)	
721 (569)	944 (745)	
850 (671)	921 (727)	
602 (473)	781 (617)	
200 (188)	139 (131)	
247 (232)	149 (140)	
276 (258)	253 (238)	
374 (252)	272 (255)	
262 (246)	226 (211)	
	157 195 223 193 232 558 (441) 682 (539) 721 (569) 850 (671) 602 (473) 200 (188) 247 (232) 276 (258) 374 (252)	

Figures in brackets are values corrected for moisture content equivalent to the unaltered rock.

Table 2. Rd values of anaerobic conditions with rock only (ml g⁻¹)

		, , , , ,	
Cs conc. (ppm)	1 day equilibration	8 days equilibration	
Unchanged rock		,	
1	213	141	
0.1	319	190	
0.01	293	213	
0.001	337	218	
0.0001	448	218	
Dry sterilised rock			
1	710 (560)	481 (379)	
0.1	1136 (896)	735 (581)	
0.01	1194 (943)	837 (662)	
0.001	1182 (934)	817 (645)	
0.0001	992 (784)	639 (505)	
Wet sterilised rock			
1	172 (162)	139 (131)	
0.1	277 (260)	177 (165)	
0.01	334 (213)	226 (212)	
0.001	276 (258)	235 (221)	
0.0001	327 (307)	199 (187)	

Figures in brackets are values corrected for moisture content equivalent to the unaltered rock.

By comparing table 1 with 2 and table 3 with 4 it is seen that the ¹³⁷Cs sorption values onto rock material are similar in the absence of introduced microbes for both aerobic and anaerobic conditions with a slight increase in the anaerobic results. However, high concentrations of Cs give lower sorption values. Unchanged and wet sterilised rock material have similar values but the dry sterilised rock has the highest sorption capacity even when corrected for the loss of moisture (tables 1 and 2).

Consistent results are produced when SRB are added under aerobic conditions, where SRB activity is not favoured, with little change when equilibration time or age of culture is varied (table 3). Values are again high when the rock has been dry sterilised as seen before in table 1. Under anaerobic conditions (table 4), which favour SRB

Table 3. Rd values for aerobic conditions with rock and KSRB (ml g⁻¹)

Cs conc. (ppm)	1 day equilibration Age of KSRB (days)		8 days equilibration Age of KSRB (days)	
	1	14	1	14
Unchanged rock				
1	162	236	174	172
0.1	235	473	225	235
0.01	265	332	256	269
0.001	197	305	245	233
0.0001	279	280	260	233
Dry sterilised roc	k			
1	623 (492)	558 (441)	584 (395)	578 (452)
0.1	978 (772)	1164 (920)	863 (715)	935 (745)
0.01	1091 (862)	997 (787)	1048 (817)	953 (743)
0.001	958 (757)	2369 (1864)	949 (748)	944 (754)
0.0001	796 (629)	587 (461)	732 (584)	963 (752)
Wet sterilised roc	k			
1	231 (216)	1669 (1569)	199 (187)	175 (164)
0.1	339 (318)	248 (233)	203 (191)	224 (210)
0.01	428 (402)	316 (296)	225 (211)	245 (229)
0.001	485 (456)	286 (268)	230 (216)	265 (249)
0.0001	272 (255)	316 (296)	245 (229)	236 (222)

Figures in brackets are values corrected for moisture content equivalent to the unaltered rock.

Table 4. Rd values for anaerobic conditions with rock and KSRB (ml g⁻1)

Cs conc. (ppm)	1 day equilibration Age of KSRB (days)		8 days equilibration Age of KSRB (days)	
	1	14	1	14
Unchanged rock				
1	30	28	26	29
0.1	505	439	483	498
0.01	157	226	244	230
0.001	72	87	105	104
0.0001	829	1075	1097	1293
Dry sterilised roo	c k			
1	139 (110)	116 (91)	108 (85)	113 (89)
0.1	2822 (2229)	2197 (1736)	2197 (1736)	2208 (1744)
0.01	944 (745)	987 (780)	1080 (853)	1346 (1063)
0.001	441 (348)	433 (341)	513 (404)	529 (417)
0.0001	3866 (3055)	4537 (3583)	4316 (3409)	4793 (3786)
Wet sterilised ro	c k			
1	24 (23)	22 (21)	23 (22)	28.(26)
0.1	464 (436)	367 (344)	441 (415)	564 (550)
0.01	255 (239)	211 (198)	228 (214)	313 (294)
0.001	112 (104)	92 (87)	96 (90)	134.(126)
0.0001	1141 (1072)	773 (726)	1313 (1233)	1596 (1500)

Figures in brackets are values corrected for moisture content equivalent to the unaltered rock.

activity, the sorption values are rather erratic, some being higher than the aerobic values and some lower. This is not due to equilibration time nor age of culture. The values for the unchanged and the wet sterilised rock are similar (whether corrected for moisture content or not) whilst those from the dry sterilized rock are much higher. It appears that the presence of SRB in conditions where their activity is favoured has complex effects on ¹³⁷Cs sorption. There is little change in Rd values for the unchanged rock even after 8 days equilibration (tables 1 and 3). Values for the dry sterilised rock are also similar

except for a few anomalogous values in table 3. Values for the wet sterilised rock under aerobic conditions are similar.

From tables 2 and 4 it is obvious that the presence of microbes in anaerobic conditions causes changes in the Rd values. Some values are higher than in the 'rock only' experiments whereas other values are lower. Again this variation is not related to KSRB age nor to equilibration time but rather on Cs concentration.

Discussion

The Rd values obtained suggest that Cs and, under anaerobic conditions, microbes are sorbed onto the rock material. Addition of KSRB to the rock material gives variable Rds when compared to the rock only results. Lower Rds suggest competition for the rock sorption sites between the microbes and Cs ions. Microorganisms are larger than Cs ions and one microbe could cover many sorption sites normally available for Cs, thus confining Cs more to the aqueous phase with a resulting lower Rd. Lower Rds may be related to the formation and action of biofilms 7, 15. Microbial activity increases after adhesion⁸. The metabolic activity of microbes sorbed onto clays has been studied 20. Once microbes sorb onto a surface in vivo they produce, amongst other things (e.g. pili⁵), a glycocalyx of extracellular polysaccharides 6 which eventually form a biofilm enabling permanent microbial adhesion and which also maintains an optimum environment for growth. Under stress conditions, glycocalyx formation is promoted 6, 10, 11. The low nutrient conditions given in the experiment may have acted similarly as a stressed environment. It is possible that the slightly varying KSRB inoculation sizes have resulted in varying amounts of biofilm. KSRB as gramnegative bacteria bind metals from solution into their walls 1. This would lower the amount of Cs in the aqueous solution with resulting variations in Rds dependent on the amount of biofilm formation.

Conclusions

Several broad findings can be extrapolated from the data presented. Since wet sterilised and unaltered rock show little difference the microbes present in the rock naturally do not appear to influence sorption. However the presence of KSRB in anaerobic conditions influences irregularly the partition of Cs, possibly in an active process. Variations and fluctuations in Rd do not occur in aerobic conditions. The age of the cultures as well as a prolonged equilibration time do not influence sorption characteristics. Dry sterilised rock with or without SRB has a higher sorption capacity than unaltered or wet sterilised rock, probably as a result of changes in the montmorillonite layer structure.

Clearly, these findings have important implications for anaerobic batch sorption experiments routinely used for assessing the sorption capacity of potential host rocks for radioactive waste repositories. Microbial contamination will lead to anomalous results, this has also to be taken into consideration for field tests where migration of a radioactive tracer is studied.

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- 1 Beveridge, T. J., and Fyfe, W. S., Metal fixation by bacterial cell walls. Can. J. Earth Sci. 22 (1985) 1893-1898.
- 2 Brookins, D. G., Geochemical Aspects of Radioactive Waste Disposal. Springer Verlag, New York 1984.
- 3 Chapman, N. A., and McKinley, I. G., The Geological Disposal of Nuclear Waste. Wiley, Chichester 1987.
- 4 Christofi, N., West, J. M., and Philp, J.-C., The geomicrobiology of European mines relevant to nuclear waste disposal. Report of British Geological Survey Fluid Processes Research Group. FLPU 85-1 (1985).
- 5 Corpe, W. A., Microbial surface components involved in adsorption of microorganisms onto surfaces, in: Adsorption of Microorganisms to Surfaces, pp. 105-144. Eds G. Bitton and K. C. Marshall. John Wiley, New York 1980.
- 6 Costerton, J. W., Marrie, T. J., and Cheng, K.-J., Phenomena of bacterial adhesion, in: Bacterial Adhesion, pp. 3-44. Eds D. C. Savage and M. Fletcher. Plenum Press, New York 1985.
- 7 Daniels, S. L., Mechanisms involved in sorption of microorganisms to solid surfaces, in: Adsorption of Microorganisms to Surfaces, pp. 7– 58. Eds G. Bitton and K. C. Marshall. John Wiley, New York 1980.
- 8 Fletcher, M., Effect of solid surfaces on the activity of attached bacteria, in: Bacterial Adhesion, pp. 339-362. Eds D. C. Sauvage and M. Fletcher. Plenum Press, New York 1985.
- 9 Gale, N. L., The role of algae and other microorganisms in metal detoxification and environmental clean-up. Biotech. Bioeng. Symp. 16 (1986) 171-180.
- 10 Govan, J. R. W., Mucoid strains of *Pseudomonas aeruginosa*: The influence of culture medium on the stability of mucus production. J. med. Microbiol. 18 (1975) 513-522.
- 11 Govan, J. R. W., and Fyfe, J. A. M., Mucoid *Pseudomonas aeruginosa* and cystic fibrosis: Resistance of the mucoid form to carbenicillin, flucloacillin, tobramycin and the isolation of mucoid variants in vitro. J. Antimicrob. Chemotherapy 4 (1978) 233-240.
- 12 KBS, Final storage of spent nuclear fuel -KBS3. 4 volumes. Swedish Nuclear Fuel Supply Co. (SKBFG/KBS) Stockholm (1983).

- 13 Lorenz, M. G., and Krumbein, W. E., Uranium mobilisation from low-grade ore by cyanobacteria. Appl. Microbiol. Biotechn. 21 (1985) 374-377.
- 14 Mann, H., and Fyfe, W. S., Uranium uptake by algae: experimental and natural environments. Can. J. Earth Sciences 22 (1985) 1899– 1903.
- 15 Marshall, K. C., Mechanisms of bacterial adheison at solid-water interfaces, in: Bacterial Adhesion, pp. 133-162. Eds D. C. Savage and M. Fletcher. Plenum Press, New York 1985.
- and M. Fletcher. Plenum Press, New York 1985.

 16 Mowll, J. L., and Gadd, G. M., Cadmium uptake by Aureobasidium pullulans. J. gen. Microbiol. 130 (1984) 279-284.
- 17 NAGRA, Projekt Gewähr 1985. Vols 1-8 NTB 85-01 to 85-08 (German) and English Summary; NAGRA NTB 85-09, Baden, Switzerland 1985.
- 18 Poole, E. G., and Kelk, B., Calcium montmorillonite (Fullers' Earth) in the Lower Greensand of the Baulking area, Berkshire. Report of the Institute of Geological Sciences No. 71/4 (1971).
- Postgate, J. R., The Sulphate Reducing Bacteria. Cambridge University Press, Cambridge 1984.
- 20 Stotzky, G., and Rem, L. T., Influence of clay minerals on microorganisms. 1. Montmorillonite and kaolinite on bacteria. Can. J. Microbiol. 12 (1966) 547-563.
- 21 Tobin, J. M., Cooper, D. G., and Neufeld, R. J., Uptake of metal ions by *Rhizopus arrhizus* biomass. Appl. envir. Microbiol. 47 (1984) 821– 824.
- 22 West, J. M., McKinley, I. G., and Chapman, N. A., Microbes in deep geological systems and their possible influence on radioactive waste disposal. Radiocat. Waste Manag. nucl. Fuel Cycle 3 (1982) 1-15.
- 23 West, J. M., Christofi, N., and McKinley, I. G., An overview of recent microbiological research relevant to the geological disposal of nuclear waste. Radioact. Waste Manag. Nucl. Fuel Cycle 6 (1985) 79-95.
- 24 West, J. M., and Arme, S. C., Tolerances of microorganisms to extreme environmental conditions. Report of British Geological Survey Fluid Processes Research Group FLPU 85-14 (1985).
- 25 West, J. M., Abbott, M. A. W., Rowe, E. J., Microbial activity in nutrient depleted materials. Report of British Geological Survey Fluid Processes Research Group FLPU 86-8 (1986).
- 26 West, J. M., Haigh, D. C., Hooker, P. J., and Rowe, E. J., Radionuclide sorption onto Fullers' Earth (calcium montmorillonite) the influence of sulphate reducing bacteria. Report of British Geological Survey Fluid Processes Research Group FLPU 87-4 (1987).
- 27 White, C., and Gadd, G. M., The uptake and cellular distribution of zinc in Saccharomyces cerevisia. J. gen. Microbiol. 133 (1987) 727– 737.
- 28 Zajic, J. E., Microbial Biogeochemistry. Academic Press, New York 1969.

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